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Post-Treatment Circulating Free HPV DNA As a Marker of Treatment Outcome in Patients with HPV-Related Propharyngeal Cancer After Radio(chemo)therapy

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Abstract

Circulating cell-free HPV16 DNA (cfHPV DNA) assessed after treatment completion was tested as a prognostic marker of treatment failure in patients with HPVrelated oropharyngeal cancer (OPC) treated with radiotherapy alone (RT) or radiotherapy combined with chemotherapy (CHRT). Fifty-five patients with OPC in whom cfHPV DNA was found prior to RT/CHRT were included into the study. cfHPV DNA was subsequently assessed after treatment completion. cfHPV DNA remission (cfHPVrem) was defined as the absence of cfHPV DNA after treatment completion. The association between cfHPVrem and the risk of local, nodal or distant failure was calculated. Survival curves were calculated according to the Kaplan-Meier technique and compared according to cfHPVrem after treatment completion with the log-rank test. Thirteen (24%) patients presented no cfHPVrem after treatment completion. For these patients, the risk of nodal failure was significantly higher (HR=9.38, 95% CI: 1.63-53.97, p=0.009) than for patients with cfHPVrem. The probability of dissemination was comparable in patients with cfHPVrem and with no cfHPVrem (p=0.95). For patients with HPV related OPC, who has no cfHPVrem after RT/CHRT completion stricter follow up is proposed due to higher risk of nodal failure.

Keywords: Circulating-free HPV DNA; Head and neck cancer; Radichemotherapy; Liquid biopsy

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Introduction

Locoregional relapse is the main reason of treatment failure in patients with head and neck squamous cell carcinoma (HNSCC) who underwent radiotherapy (RT) alone or combined with chemotherapy (CHT). Despite treatment intensification for locoregionally advanced HNSCC, approximately half of the treated population returns with recurrent or refractory disease and for these patients, the 1-year survival rate ranges from 5% to 33% with a median OS of 6 months to 9 months [1,2]. Early detection of treatment failure is essential for successful salvage [3] but challenging due to the lack of reliable markers and low sensitivity of imaging methods directly after treatment [4,5]. A causal association between human papillomavirus (HPV) and a subset of HNSCC especially oropharyngeal cancer (OPC) has changed a previous dogma as to epidemiology, pathology and clinical course of HNSCC. Recent studies showed that circulating

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Materials and Methods

Newly diagnosed patients with OPC who had been admitted to the Radiation Oncology Department at Maria Sklodowska-Curie Memorial Cancer Center and Institute of Oncology in Gliwice between September 2012 and May 2016 were eligible for inclusion to the study. Patients with known metastatic disease or immune suppression were not eligible. At the time of study recruitment (presentation to the institution), informed consent was obtained from all the patients. The project was approved by the Bioethics Committee at the Maria Sklodowska-Curie Memorial Cancer Centre and Institute of Oncology. The study conforms with the Code of Ethics of the World Medical Association. Plasma of 179 patients was obtained for cfHPV DNA assessment prior to any treatment. Circulating free HPV DNA was found in 55 (31%) patients and subsequently serial cfHPV DNA blood samples were taken after induction CHT (indCHT) for those who underwent indCHT and directly after the treatment completion in all the patients. HPV-related tumor etiology was confirmed in tissue samples in 47 (84%) cases from this group. For 8 (16%) patients tissue samples were not available or not informative. The group consisted of 28 (52%) men and 27 (48%) women at the mean age of 55 years (range: 30 years to 75 years). In 5 (9%) patients metastatic neck lymph nodes had been dissected as a diagnostic procedure prior to the presentation at the Radiation Oncology Department. All the patients were treated definitively with RT or RT combined with CHT in 6 (11%) and 49 (89%) cases respectively. Induction chemotherapy was performed in 30 (53%) patients. It was followed with RT alone in 8 (14%) or with radiochemotherapy (CHRT) in 22 (39%) cases. In 2 (5%) patients after indCHT palliative RT was given due to progressive disease. Clinical data in details is presented in **Table 1**.

Plasma samples collection and DNA extraction

Peripheral blood (12 ml) was collected into $\rm K_3EDTA$ tubes (Becton-Dickinson, New Jersey, Franklin Lakes, USA). Plasma was immediately (up to 1 hour) separated by double centrifugation at $300 \times \rm g$ and $1000 \times \rm g$, both at 4°C for 10 minutes. Aliquots of the plasma were frozen at -80°C. DNA was extracted (according to the manufacturer's instructions) from 1 ml of plasma by the Genomic Mini AX Body Fluids kit (A&A Biotechnology, Gdynia, Poland) and dissolved in 20 μ l of the elution buffer.

Analysis of the total cfDNA and HPV16 in blood plasma

For the total cfDNA (circulating cell-free DNA) measurement amplification of TERT (human telomerase reverse transcriptase) was used as described previously [11]. Shortly, each measurement consisted of a standard curve, negative control and a sample. For the construction of the standard curve, we used a control genomic DNA. The concentration of the total cfDNA was between 0.118 ng/µl and 1.130 ng/µl. The concentration of the total cfDNA did not influence the cfHPV DNA detection as we presented before [11-16]. For HPV16 detection primers and probe sequence were designed for E6/7 region. PCR was done in a final volume of 30 µl using the TaqMan Genotyping Master Mix (Applied Biosystems, CA, Foster City, USA). If HPV16 was found, the confirmation of its presence was done on the second independent DNA isolation. Complete remission of cfHPV DNA (cfHPVrem) was defined as not detectable cfHPV DNA in plasma after the treatment.

Tumour samples: Detection of HPV and confirmation of its biological activity in the tumour tissue

DNA was isolated from formalin-fixed paraffin-embedded (FFPE) tumour samples using the mSample Preparation System DNA (Abbott Laboratories, Abbott Park, Illinois, USA) (N=33). Detection of HPV in FFEP tumour samples was made using a Real-time high-risk HPV test (Abbott Molecular, Abbott Park, Illinois, USA). The reactions contained 50 ng of extracted DNA. Samples were considered to be positive when the $C_{\rm t}$ value, which means the number of cycles of which the fluorescence exceeded the threshold, was less or equal to the cut-off value of 35. Samples for which the β -globin $C_{\rm t}$ value was >35, were considered to be non-informative.

Statistical Analysis

Due to the treatment method patients were divided into these who underwent indCHT and those who did not. Locoregional recurrence free survival was defined as a time with no evidence of the disease in primary tumor site (local recurrence free survival) and regional lymph nodes (nodal recurrence free survival). Metastases free survival was defined as a time with no evidence of distant spread of the disease. The rate of treatment failure according to cfHPVrem after the treatment completion was calculated and compared with Fisher's exact test. Survival curves were calculated according to the Kaplan-Meier technique and compared according to cfHPVrem after the treatment completion with the log-rank test.

Results

Treatment outcome according to cfHPV DNA status after treatment completion

Among 30 (53%) patients who underwent indCHT, six patients had 1 course, four patients had 2 courses, sixteen patients had 3 courses and four patients had 4 courses of indCHT (Table 1).

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Table 1 Patients characteristic according to clinical factors, treatment and treatment results according to cfHPV DNA status after indCHT and after treatment completion.

Case				indCHT	Radiotherapy	cfHPVrem after: indCHT/	Treatment	Follow-up
number	Sex	HPV in tissue	TN	(no of courses)	(Gy)	treatment completion	results	(months)
1	М	Positive	T3N3	4	CHRT(70)	0/0	NED	46
2	М	Positive	T4N1	0	CHRT(70)	-/1	NED	42
3	М	Positive	T2N2	3	CHRT(70)	0/0	NED	47
4	F	Positive	T3N3	3	RT(20)	1/1	LRF	6
5	М	Positive	T3N3	3	CHRT(70)	0/0	NED	46
6	F	Positive	T4N1	0	CHRT(70)	-/1	NED	9
7	F	N/A	T3N1	0	CHRT(70)	-/0	NED	44
8	М	Positive	T3N2	0	RT (72)	-/0	NED	27
9	F	Positive	T4N2	1	RT(60)	0/0	NED	41
10	М	Positive	T4N1	0	RT (60)	-/0	NED	37
11	М	Positive	T3N3	4	CHRT(70)	0/0	NED	33
12	М	Positive	T1N3	1	CHRT(66)	0/0	NED	36
13	М	N/A	T2N3	4	RT (72)	0/0	DM	6
14	F	Positive	T3N2	3	RT(70)	0/0	NED	30
15	F	Positive	T4N2	3	RT (20)	1/1	LRF	23
16	F	Positive	T4N2	3	CHRT(70)	0/0	NED	34
17	F	Positive	T4N2	3	CHRT(70)	0/0	RF	34
18	F	Positive	T4N2	2	CHRT(70)	0/0	NED	22
19	М	Positive	T2N3	3	CHRT(70)	1/0	NED	31
20	М	Positive	T2N2	0	CHRT(70)	-/0	NED	35
21	F	N/A	T2N2	0	RT (66)	-/0	NED	35
22	F	N/A	T2N2	0	CHRT(70)	-/0	NED	16
23	M	Positive	T4N2	0	CHRT(70)	-/0	NED	5
24	M	Positive	T3N2	4	CHRT(70)	1/0	DM	24
25	М	N/A	T3N2	3	CHRT(70)	1/0	NED	28
26	F	Positive	T4	1	CHRT(70)	0/0	LF	18
27	M	Positive	T4N3	1	RT(70)	1/1	RF,DM	23
28	M	Positive	T3N2	2	CHRT(70)	0/0	NED	23
29	F	Positive	T3N3	3	CHRT(70)	1/1	RF	19
30	F	Positive	T1pN2	0	CHRT(70)	-/0	NED	22
31	M	N/A	T3N1	3	CHRT(70)	1/0	NED	21
32	F	Positive	T1pN3	0	CHRT(70)	-/1	NED	22
33	F	N/A	T2pN2	0	RT(66)	-/0	DM	17
34	F	Positive	T1N2	0	RT(70)	-/0	NED	22
35	F	Positive	T1N2	2	CHRT(70)	1/1	NED	13
36	F	Positive	T4N0	0	CHRT(70)	0/1	NED	13
37	M	N/A	T2N3	3	CHRT(70)	0/0	NED	11
38	M	Positive	T3N2	3	CHRT(70)	1/1	NED	12
39	M	Positive	T3N2	0	CHRT(70)	-/1	NED	13
40	M	Positive	T2N2	0	CHRT(70)	-/0	NED	12
41	M	Positive	T2N2	0	CHRT(70)	-/0	NED	14
42	F	N/A	T1N2	1	CHRT(70)	0/0	NED	11
43	F	Positive	T3N2	2	CHRT(70)	0/0	NED	9
44	F	Positive	T4N1	0	CHRT(70)	-/1	NED	9
45	M	Positive	T1pN2	0	CHRT(70)	-/0	NED	6
46	M	Positive	T2N0	0	RT(70)	-/0	NED	11
47	M	Positive	T2N2	3	CHRT(70)	0/0	NED	7
48	M	Positive	T2N2	0	RT(70)	-/1	NED	10
49	F	Positive	T2N2	3	RT(70)	1/0	NED	4
50	F	Positive	T3N3	3	RT(70)	0/0	NED	5
51	F	Positive	T1N2	0	CHRT(70)	-/0	NED	6
52	М	Positive	T1pN2	0	CHRT(70)	-/0	NED	4

Case numbe	Sex	HPV in tissue	TN	indCHT (no of courses)	Radiotherapy (Gy)	cfHPVrem after: indCHT/ treatment completion	Treatment results	Follow-up (months)
53	M	N/A	T1N2	1	CHRT(70)	1/0	NED	4
54	F	Positive	T1N2	0	CHRT(70)	-/0	NED	2
55	F	Positive	T2N2	0	CHRT(70)	-/0	NED	3

N/A: data not available; indCHT: Induction Chemotherapy; RT: Radiotherapy; CHRT: Radiochemotherapy; 0: cfHPVrem; 1: detectable cfHPV DNA; NED: No evidence of disease; RF: Regional Failure; LF: Local Failure, LRF: Locoregional Failure, DM: Distant Metastases

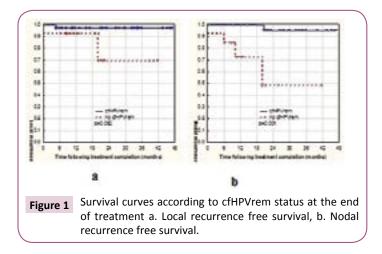
Less than 3 courses of indCHT were given due to poor toleration in five cases, or total or almost total disease remission after one or two courses in the remaining five patients. The number of courses of indCHT did not influence the rate of cfHPVrem neither after indCHT, nor after the treatment completion. cfHPVrem after the treatment completion was found in 8 (80%) and 16 (80%) patients who received 1 or 2 and 3 or 4 courses of indCHT respectively (p=1.0). Twelve patients out of 30 (36.5%) still had virus in the blood (no cfHPVrem) after indCHT, (patients 4, 15, 19, 24, 25, 27, 29, 31, 35, 38, 49, 53). Six patients from this group had subsequently no cfHPVrem after the treatment completion (patients 4, 15, 27, 29, 35, 38). Treatment failure was finally observed in 5 patients (patients 4, 15, 24, 27, 29) what is 42% of those who had no cfHPVrem after indCHT. Ultimately treatment failure was found in four patients (4, 15, 27, 29) out of six (67%) who had no cfHPVrem after the treatment completion. The risk of treatment failure in patients after indCHT who had no cfHPVrem after the induction was higher than in the others and significantly higher if no cfHPVrem was still observed after the treatment completion (p=0.029).

Among 25 patients who underwent RT or CHRT without indCHT in 18 (72%) cfHPVrem was found after the treatment completion and 7 patients had no cfHPVrem at that moment (patients 2, 6, 33, 36, 39, 44, 48). Only 1 patient from this group finally failed (patient 33).

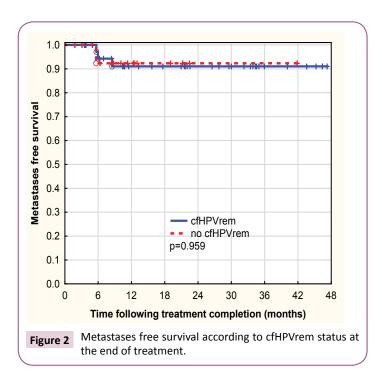
Considering together patients with indCHT and without it, in 13 (24%) patients no cfHPVrem was found after the treatment completion. The risk of subsequent failure was lower in patients with cfHPVrem after the treatment completion (12%) compared to the patients with no cfHPVrem (31%). The distribution of failures (local, nodal and distant) is presented in **Table 2**. Local failure was found in three patients. Local recurrence free survival was higher in patients with no cfHPVrem at the end of the treatment (p=0.062) (Figure 1a). Patient 4 was not cured remaining with bulky local and regional disease. Local recurrence of disease was also found in patient 26 after 5 months of complete disease remission and in patient 15, simultaneously with nodal recurrence, after 22 months of complete disease remission. Nodal failure was found in patients 4 and 15 (simultaneously with local failure mentioned above) and in patients 17, 27 (who

Table 2 The distribution of failure (local, nodal and distant) according to cfHPV status.

Variables	Local failure*	Nodal failure*	Distant failure*	Cured			
no cfHPVrem	2 (66.7%)	4 (80.0%)	1 (25.0%)	9 (19.6%)			
cfHPVrem	1 (33.3%)	1 (20.0%)	3 (75.0%)	37 (80.4%)			
р	0.122	0.01	1.0				
*separately calculated patients with more than one site of failure (patients 4, 15, 27)							



simultaneously developed metastases to the lung) and 29, after 21 months, 6 and 10 months of complete disease remission respectively. In one case, metastatic squamous cell carcinoma after salvage lymphadenectomy has been confirmed, in other two cases metastatic cancer cells were found in the fine needle aspiration. One patient denied histopathological verification of lymph nodes with FDG uptake and also the further treatment. Patients with no cfHPVrem at the end of the treatment had over twenty times higher risk of nodal failure (HR=21.88, 95% CI: 2.33-205.19, p=0.0069) (Figure 1b). In patients 13, 24, 27 (with simultaneous nodal failure) and 33 PET scanning revealed metastatic disease in lung (patient 24, 27, 33) and liver (patient 13) 6 months and 8 months after treatment completion respectively. Patients with no cfHPVrem at the end of the treatment had similar probability of dissemination as these with cfHPVrem (p=0.95) (Figure 2). Second primary (SP) was found in 3 (5%) patients. It appeared in patient 23 after 7 months (gastric cancer), in patient 9 after 12 months (lung cancer), and in patient 2 after 20 months (pancreatic cancer) after the treatment. cfHPVrem was found in these patients.



Discussion

Results of our study indicate that for the patients with HPV related OPC in whom cfHPV DNA is present prior to RT/CHRT, post-treatment cfHPV DNA status has a prognostic value. Viral DNA has been disappearing quickly in the blood of most patients during treatment but not in all cases. cfHPV DNA was still detectable in almost one fourth of all patients after the treatment completion. In this group with the exception of one patient who was not cured and one with residual neck mass who was finally cured all the others presented no evidence of disease at that moment. However, despite of total clinical clearance of disease, higher risk of the treatment failure could be expected in patients with no cfHPVrem at the end of the treatment. Thirtyone per cent of patients with virus detectable at the end of treatment (no cfHPVrem) ultimately failed, while among these with cfHPVrem-only in 12% failed. Among those who underwent indCHT (more intense therapy), the treatment failure was observed even in 67% of patients without cfHPVrem after the treatment completion. Our results indicate that persisting HPV DNA after the treatment completion may help to predict rather locoregional failure, especially nodal, but not distant spread of disease. Patients without cfHPVrem after treatment presented over 20 times higher risk of nodal failure than the patients with cfHPVrem while not difference was found in the risk of metastatic disease between both groups. Ahn et al. presented similarly adverse results of persistent HPV DNA in serum of patients after the treatment. Patients with posttreatment plasma HPV-positive status demonstrated a significant association with increased risk of disease recurrence after adjusting for other risk factors like alcohol use, smoking status, and N classification. In their study, among 5 patients with detectable serum HPV DNA after treatment 4 (75%) developed subsequently recurrence while only 2 from 30 patients (13%) who did not have HPV detected in posttreatment plasma sample [17]. Cao et al. followed small group of 14 patients who presented pretreatment detectable plasma HPV DNA in serial plasma samples (weekly or every other week) during the course concurrent chemoradiotherapy. There was a gradual decline in HPV DNA during the therapy which, disappeared at all at the end of it. There were no obvious differences in the rate of HPV DNA decline between the patients with eventual tumour relapse and those without it. Four of patients however eventually relapsed: one locoregionally and three distantly in the lungs [6]. It shows, similarly as in our study, that HPV DNA status at the end of the treatment may not be a good predictor of distant spread of the disease. Most data on the role of viral cfDNA as a prognostic marker comes from studies reporting DNA of Epstein-Barr virus (EBV) in patients who underwent treatment due to nasopharyngeal carcinoma (NPC). Circulating EBV DNA has been tested as a marker for NPC since 1998 [18,19] and correlates positively with disease stage and exhibits prognostic value in patients with NPC [20]. The drop in viral DNA in the blood during RT/CHRT was accompanied by remission of NPC as a response to treatment [21]. Patients with more rapid clearance of EBV DNA from the circulation responded better to treatment and had a better survival probability [22,23]. Due to high sensitivity and specificity [24,25], plasma EBV DNA has been adopted as a marker for clinical management of patients with NPC [26]. It was suggested that high circulating plasma EBV DNA load assessed 6 weeks post-primary treatment may predict a subsequent relapse [27-29]. For patients treated due to NPC persisting, increased, post-treatment viral DNA load or a subsequent rise after a previous drop seems to be an indication for PET/CT scanning for high risk of relapse [30]. Campitelli et al. showed an analysis of sequential serum specimens in patients with cervical cancer. The concentration of cfHPV preceded radiological relapse in one case and was associated with an 8-mm liver metastasis in the other one. The authors hypothesized that tumour DNA might be more easily released from tumour tissue corresponding to a relapse or metastasis than to a primary lesion. They concluded that during follow-up, cfHPV DNA concentration could be a highly specific surrogate marker for minimal residual disease or subclinical relapse [31]. In another study, in patients treated due to cervical cancer, plasma cfHPV DNA became undetectable after the treatment in those who had complete response and showed no evidence of disease after the treatment. Contrary to that, cfHPV DNA was still present after the treatment and during the followup in those who had persistent disease after the treatment or presented recurrence. Authors concluded that plasma cfHPV DNA might be a valuable noninvasive marker for monitoring the therapeutic response and disease progression in patients treated due to cervical cancer [32]. Some limitations may restrict conclusion that could be drawn from this study. Only patients with HPV DNA in the blood prior to the treatment were followed with the subsequent assessment of HPV DNA. It is likely that more HPV-positive patients release HPV DNA to the blood but had not been caught at the moment of our assessment prior to the treatment. In further research, all the patients with HPV-related tumorus (based on a tissue sample) should be followed with HPV DNA assessment to recognize "HPV DNA plasma windows".

Our group consisted of patients with OPC exclusively and the usefulness of HPV DNA assessment in HPV-related tumours of other sites in head and neck region has not been explored yet.

Conclusion

In summary, for patients with HPV-related OPC, cfHPV DNA seems to be a marker for active cancer. If cfHPV DNA is still present in

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the blood after the treatment completion it may indicate patients with higher risk of nodal failure. For such patients, stricter followup should be considered. Further research is needed however to confirm our results in a larger group of patients.

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